

SPECIAL ISSUE ARTICLE

Bioactive glass and glass-ceramic orbital implants

Francesco Baino¹  | Enrica Verné¹ | Elisa Fiume¹ | Oscar Peitl² |
Edgar D. Zanotto² | Simone M. Brandão³ | Silvana A. Schellini⁴

¹Department of Applied Science and Technology (DISAT), Institute of Materials Physics and Engineering, Politecnico di Torino, Torino, Italy

²Department of Materials Engineering (DEMa), Center for Research, Technology and Education in Vitreous Materials (CeRTEV), Federal University of Sao Carlos (UFSCar), Sao Carlos, Sao Paulo, Brazil

³Department of Medicine, Federal University of Sao Carlos (UFSCar), Sao Carlos, Sao Paulo, Brazil

⁴Department of Ophthalmology, Faculdade de Medicina de Botucatu, Universidade Estadual Paulista, Botucatu, Sao Paulo, Brazil

Correspondence

Francesco Baino, Department of Applied Science and Technology (DISAT), Institute of Materials Physics and Engineering, Politecnico di Torino, Torino, Italy
Email: francesco.baino@polito.it

Abstract

This review focuses on the applications of bioactive glasses and glass-ceramics in the field of orbital implants for ocular surgery. This use is relatively novel and less popular compared to the applications in orthopedics and dentistry for the repair of bone and teeth. Recent studies have shown the suitability of bioactive glasses and glass-ceramics in contact with soft tissues for promoting additional effects associated to the release of therapeutic inorganic ions. Specifically, the angiogenic and antibacterial actions that may be elicited by selected glass compositions are highly appealing for the development of new-generation orbital implants, since improved vascularization and antiseptic properties are the key for a higher success rate of anophthalmic socket procedures. An overall picture of existing orbital implants based on bioactive glasses is here provided, and the further potential and open challenges for future research in this field are highlighted and discussed.

KEY WORDS

bioactive glass, bioceramic, orbital implant, porous implant

1 | INTRODUCTION

The eye loss is disfiguring and the concern with redoing facial aesthetics is very old. Even though it is a difficult decision, there are some critical conditions, for example, blind and painful eyes, eyes with intraocular tumors or after severe trauma, in which the removal of the entire eye or its content is still mandatory.¹

Archeological findings have revealed that around 500 BC Egyptian and Roman people losing an eye wore painted prostheses (a clay shell worn over or under the lids) to recover the proper appearance.² Till the Middle Age, different combinations of moldable materials (eg, wool, clay) and noble metals (eg, gold, silver) were used to fabricate different kinds of ocular prosthesis; the thin metallic foil covering the

anophthalmic socket was often enameled or painted to reproduce the color of natural iris, thereby providing the patient's face with more acceptable aesthetics.³

The surgical techniques used to remove the eye in toto or its content evolved at the same time than the evolution of the orbital implants. There are descriptions of old techniques (16th century) using a strong wire transfixing the globe, which was then drawn until the eye was out of the orbit.⁴ The procedures of human eye removal were not clearly standardized until the end of the 19th century⁵ and then evolved quickly. Nowadays, there are two well-established main techniques: (a) evisceration which refers to the removal of the contents of the eye while the scleral shell and ocular muscle attachments are spared, and (b) enucleation which is a more radical measure involving the removal of the whole globe from the orbital socket. The majority of cases are suitable for evisceration, whereas enucleation is mandatory for

*Baino, Verné and Zanotto are Member of the American Ceramic Society (ACerS).

intraocular tumors (eg, retinoblastoma or melanoma), since without eye removal the malignant lesion could spread to the surrounding structures or metastasize.

After the enucleation or evisceration, implants must be placed in the orbit to restore the lost ocular volume.⁶ If the patient undergoes evisceration, the implant is usually wrapped by the own scleral shell and the extraocular muscles are left in place. When the surgical procedure is an enucleation, the implant is wrapped within a foil of donor sclera or smooth polymeric mesh to which the patient's extraocular muscles are sutured. The Tenon's capsule and the conjunctiva must be closed over the sclera and in the anterior portion of the orbital cavity in order to prevent the exposure of the implant, avoid conjunctival abrasion, “isolate” the implant from the outer environment and protect it from foreign pathogens (Figure 1A).

Since the late 1500s, Venetian glassmakers in Italy began to produce the early so-called “glass eyes,” that is, hollow spheres of blown glass that were inserted inside the empty orbital cavity.⁷ Although being fragile and needing to be worn cautiously by patients, these implants rapidly gained success and were also exported to other European countries.

At the end of the 19th century, Mules reported a detailed description of a hollow glass sphere and its placement inside the orbit to replace the loss of volume in an enucleated socket.⁵ This type of implant was the standard option in anophthalmic socket surgery until the end of the Second World War. The most important factory of glass orbital implants in the first half of the 20th century was located in Germany and was

destroyed in the 1940s⁴; since then, glass orbital implants had fallen in almost total disuse.

From 1950 to 1970, silicone or poly(methyl methacrylate) (PMMA) solid balls⁴ became the most popular materials for replacing the volume of anophthalmic socket. Both these materials, as well as the previously applied glass spheres, have a smooth surface that do not interact either with the host tissues or with the external ocular prosthesis—which is usually a painted PMMA shell mimicking the aesthetics of the contralateral eye—and hence they were named nonintegrated implants.

The anophthalmic socket reconstruction scenario dramatically changed in the 1980s with the advent of porous orbital implants.^{8–10} Porous implants allowed improving the clinical success rate and the life of the implant due to postoperative fibrovascular colonization inside the implant. While nonporous polymeric materials are typically embedded in a collagenous pseudocapsule once implanted *in vivo*, the connective tissue composed by vessels and inflammatory host cells contributing to the healing process invades the void network of the porous implant and anchors it to the orbital tissues. This is believed to minimize the risk of postoperative implant migration.¹¹ Furthermore, there is a convincing evidence that porous implants lead to better clinical outcomes compared to nonporous devices as the presence of a blood supply within the implant permits immune surveillance as well as the treatment of bacterial infection via systemic antibiotics.¹² Some reports have also suggested that fibrovascularization could promote the spontaneous healing of small exposures of the

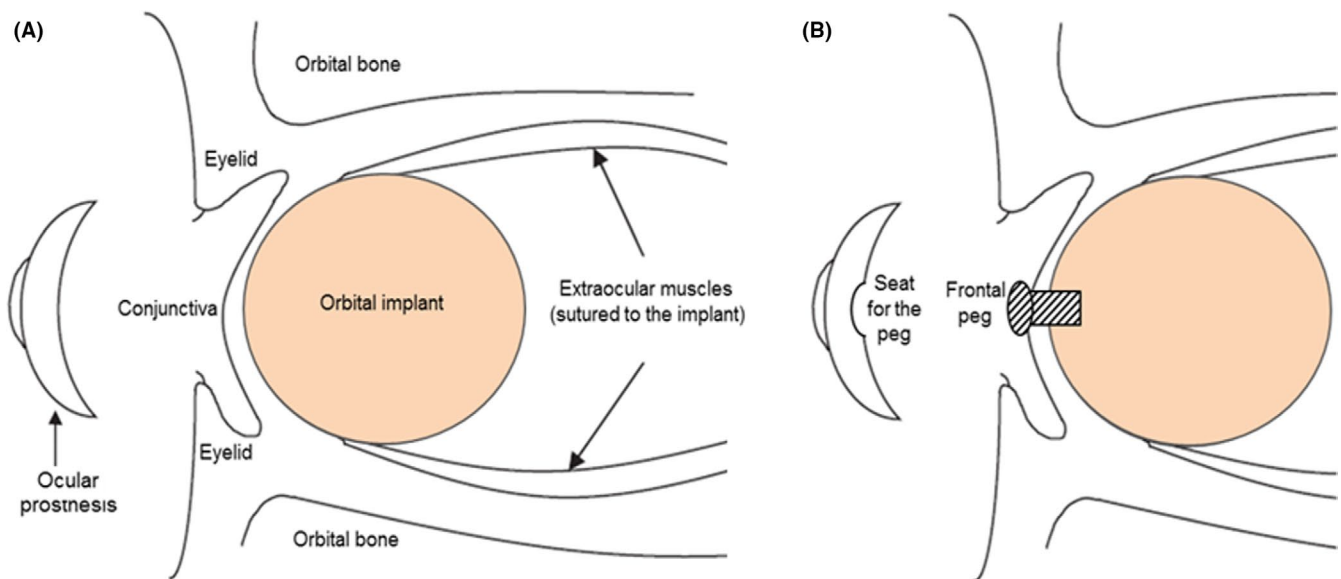


FIGURE 1 Schematic illustration showing a spherical orbital implant placed in the anophthalmic cavity after enucleation. The orbital implant can be (A) “buried” under the patient’s conjunctiva (“nonintegrated” implant) without any mechanical connection to the ocular prosthesis or (B) connected to the ocular prosthesis by a peg (“integrated” implant). Pegged implants, although allowing a wider range of movements to the ocular prosthesis, are seldom adopted nowadays due to the need for a second surgery for peg placement. Image reproduced from Ref. ⁷⁰ [Colour figure can be viewed at wileyonlinelibrary.com]

implant¹³; however, this potential advantage is still under debate.

Over the years, there was some confusion about the meaning of the term “integrated” in the context of orbital implants because for some authors “integration” means the tissue reaction inside the implant—which typically occurs in porous implants—while, according to others, integrated implants are the ones which can receive a peg system for direct coupling with the external prosthesis⁹ (Figure 1B). A first, clear attempt to solve this issue was carried out in 2002 when the scientific panel of the American Academy of Ophthalmology established that “integration” refers to the nature of fit between the external ocular prosthesis and the orbital implant. However, this controversy is still far from the end.

Porous orbital implants available on the market are made of natural or synthetic hydroxyapatite, polyethylene, or alumina and exhibit a network of open and interconnected macropores in the range of 100–500 μm .¹⁰ There is evidence that porous alumina implants lead to better clinical outcomes than both porous hydroxyapatite, mainly due to the lower surface roughness (less than 1 μm vs few micrometers¹⁴) which minimizes the risk of implant exposure, and porous polyethylene that, however, exhibits quite smooth walls.¹⁵ This observation can be explained in light of the faster fibrovascularization rate of alumina implants, which is promoted by the more favorable chemical surface of the material. In fact, cell and tissue ingrowth within porous implants is accelerated by hydrophilic surfaces like those of alumina, and discouraged by hydrophobic polymers that tend to be encapsulated in a fibrous collagenous capsule.¹⁶

In spite of all these attractive characteristics, commercial porous implants—and especially alumina implants—are not accessible to a large number of patients due to the high cost. Very interestingly, Sousa et al reported that the traditional, nonintegrated PMMA implant is still the most commonly used all over the world,¹⁷ and other studies indicate that cheap nonintegrated silicone spheres are in widespread use in developing countries.¹⁸

At present, the majority of orbital implants are spherical; however, other “styles” and formats were developed over the years. A special mention should be dedicated to the “quasi-integrated” implant, which was introduced in the 1970s and was characterized by a “lock-and-key” coupling system to better support the external ocular prosthesis and expand its range of movements.¹⁹ Specifically, this PMMA implant exhibited a grossly semispherical geometry provided with four anterior mounds that matched four corresponding depressions on the posterior surface of the ocular prosthesis.²⁰ The mounds created two perpendicular channels so that the stumps of horizontal and vertical extraocular muscles could be sutured together before being covered by the conjunctiva. It is worth pointing out that there was no interruption of conjunctival lining, but the irregular anterior surface of

the implant was used to improve translation of implant movement to external prosthesis movement. This type of implant underwent several evolutions over time and it was shown that the adoption of a conical geometry can have better contact with the extraocular muscles and maybe can be a good option to increase the external prosthesis movements along both vertical and horizontal axes.²¹

Complications related to orbital implants may happen due to inherent characteristics of the implant material. For example, the exposure rate of porous implants can be favored by a rough and stiff surface.^{22,23} Exposure in porous implants can be treated with conservative management using pharmacological treatment or salvage strategies (anterior apposition of scleral or polymeric patches), without the need for implant removal and replacement with a new one.^{24,25} Furthermore, there are other factors not directly related to the implants which can lead to postoperative complications, such as bad surgical technique or patient's systemic diseases.

2 | WHY USING BIOACTIVE GLASSES FOR MAKING ORBITAL IMPLANTS?

Despite the variety of options available on the market, nowadays an orbital implant to be considered as “ideal” does not exist. The non-negligible drawbacks of current solutions have been motivating further research in terms of both implant design and materials used.

There are multiple reasons behind the recent “resurrection” of glass as a material for orbital implants after a hiatus of about 50 years. This is a typical demonstration of how old, temporarily discarded materials can be somehow reinvented in light of new scientific and technological advances. In the 1950s, the hollow spheres of blown glass were replaced by nonporous acrylic or silicone orbital implants that were relatively light, mechanically more compatible with orbital tissues due to lower stiffness, and not prone to sudden and traumatic brittle fracture as the thin-walled glass balls might be.^{26,27}

Furthermore, today's manufacturing techniques allows glass products to be easily produced in a porous form by means of foaming, porogen removal or replication strategies,²⁸ which were not available at the time of first-generation blown glass orbital implants. Glass possesses an exceptional versatility from both compositional and technological viewpoints and can be processed at lower temperature and cost compared to other orbital implant materials, such as alumina.

In addition to the previous considerations, there is a more profound and substantial reason that differentiate these new glass-based orbital implants not only from the early glass spheres but also from all other porous and nonporous existing options: instead of using biocompatible but inert glass compositions, researchers began to investigate the suitability of

orbital implant biomaterials able to actively interact with living cells and tissues, eliciting specific biological responses.

Bioactive glasses were first developed in 1969 by Hench and coworkers, who designed the famous 45S5 composition ($45\text{SiO}_2\text{-}24.5\text{CaO-}24.5\text{Na}_2\text{O-}6\text{P}_2\text{O}_5$ wt%).²⁹ This glass, commercialized under the tradename of Bioglass[®], was found able to bond to living bone and stimulate the genes of bone cells toward paths of regeneration and self-repair³⁰; it is currently used for making many clinical products (eg, cast monoliths, micrometric particles, porous granules, injectable putty) for bone defect filling in orthopedics and dentistry.^{31,32} These bioactive glass products are capable to bond to host bone forming a tight interface and to promote the growth of new bone tissue while dissolving over time. The creation of a glass-bone bond is attributed to the formation of a nanocrystalline hydroxyapatite layer that interacts actively with the collagen fibrils of the living bone.³³ The formation of a bond *in vivo* between this surface layer and the host bone is a complex process involving protein adsorption, incorporation of collagen fibrils, adhesion of osteoprogenitor cells, cell differentiation, production, and mineralization of bone extracellular matrix.³⁴ The surface nano-hydroxyapatite layer forms following solution-mediated dissolution of the bioactive glass according to a process similar

to the corrosion of conventional glasses.³⁵ Accumulation of dissolution products causes both the chemical composition of the glass surface and the pH of the body fluids to change locally, thus providing surface sites and a pH favorable to hydroxyapatite nucleation. Once the surface hydroxyapatite layer has formed, proteins are adsorbed on it and cells can attach, differentiate, and produce new bone matrix.³⁶

Formation of a surface hydroxyapatite layer is not a goal in the field of orbital implants, but other properties of suitable biocompatible glasses can be very appealing for such application. Over the years, a number of other silicate, borosilicate and phosphosilicate glass and glass-ceramic compositions have been developed for biomedical use³⁷ and, very interestingly, some of them were found suitable for use in contact with soft tissues.³⁸ In fact, ionic dissolution products released from bioactive glasses (Figure 2) can stimulate not only osteogenesis but also angiogenesis, which is the key to accelerate wound healing and tissue regeneration.³⁹ The formation of new blood vessels is of utmost importance for ensuring the delivery of nutrients, growth factors, and oxygen, as well as for allowing stem cells to reach the injured site.

The original 45S5 Bioglass[®] was widely proved able to stimulate angiogenesis both *in vitro* and *in vivo* (rat model)

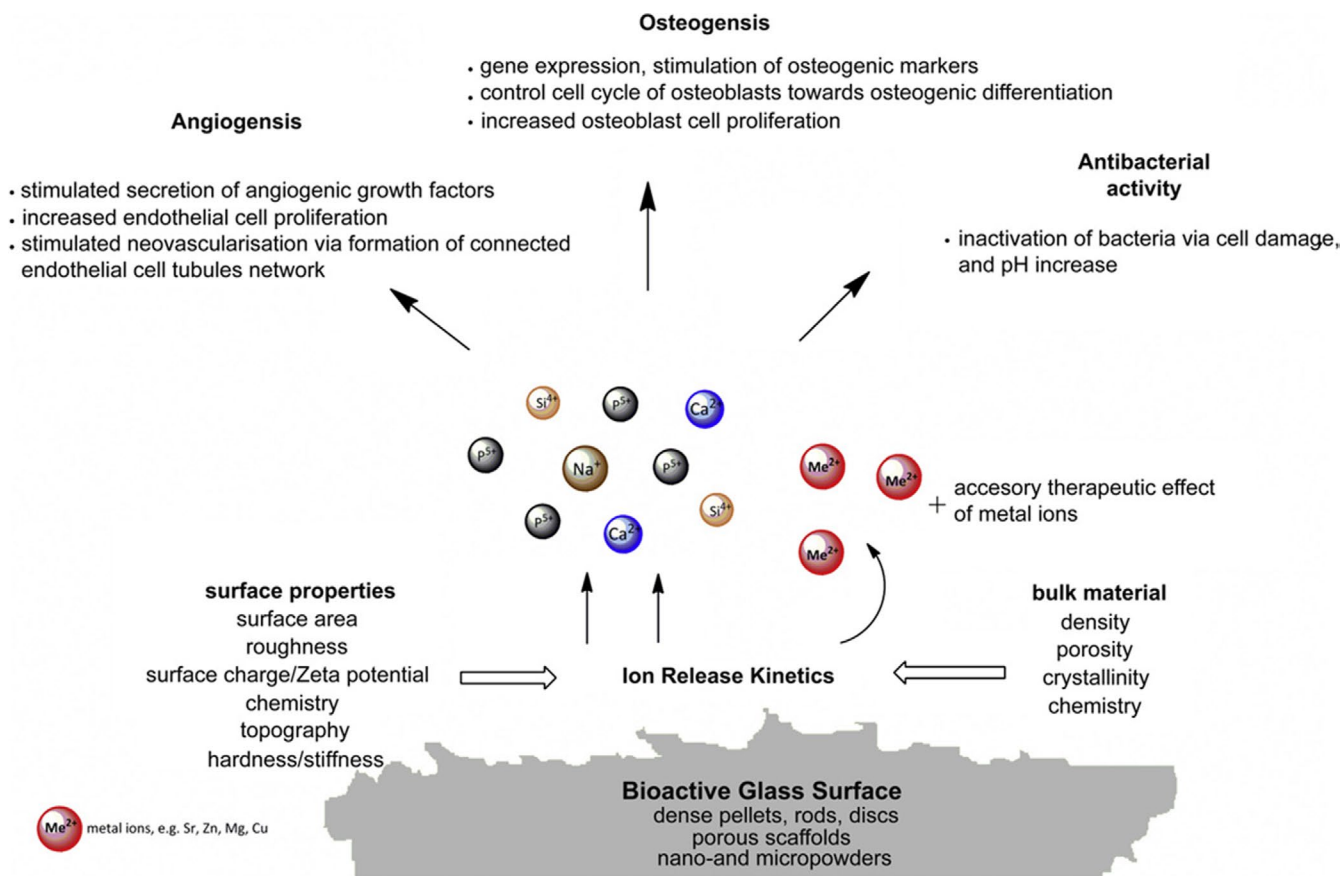


FIGURE 2 Overview of the main biological responses that can be elicited by the ionic dissolution products released from bioactive glasses once implanted *in vivo* (bone regeneration, angiogenesis, antibacterial effect). Image reproduced from Ref. ⁹⁷ [Colour figure can be viewed at wileyonlinelibrary.com]

via the release of silicate and calcium ions.⁴⁰ In fact, silicate ions can induce endothelial cell homing, polarization and migration, and sprouting of new blood vessels^{41,42}; calcium ions increase the gene expression of platelet-derived growth factor, endothelial growth factor, insulin-like growth factor-1, basic fibroblast growth factor (bFGF), and vascular endothelial growth factor (VEGF), thereby promoting the proliferation of endothelial cells.^{43,44} Aside from 45S5 composition, this property was also revealed in other bioactive glasses, such as 58S (58.2SiO₂-32.6CaO-9.2P₂O₅ wt%)⁴⁵ and 13-93B3 (56.6B₂O₃-5.5Na₂O-11.2K₂O-18.5CaO-4.6MgO-3.7P₂O₅ wt%).⁴⁶

Proangiogenic effect is highly relevant and attractive for applications in the field of orbital implants: while porous polyethylene, hydroxyapatite, and alumina act as a passive framework for fibrovascular ingrowth, the rate of which seems to be mainly dictated by surface wettability, bioactive glass implants can release ionic dissolution products that greatly stimulates angiogenesis, thus accelerating the fibrovascular reaction inside the implant.

Looking at the existing literature, bioactive glasses have been mainly (a) used to fabricate porous or monolithic glass-ceramic products, or (b) embedded as bioactive inclusions within a porous polymeric matrix to create a composite, or else (c) applied as a coating on a non-resorbable porous framework.⁴⁷⁻⁵¹ These strategies were also pursued in the field of orbital implants, as described in the next sections. Specifically, the approaches (b) and (c) are motivated by the need for retaining a permanent skeleton that supports the periocular tissues over the patient's lifetime as the bioactive glass slowly dissolves.

3 | IN VIVO STUDIES AND CLINICAL APPLICATIONS

3.1 | Early trials

After being fascinated by the interaction between bioactive glasses and living tissues, a group of Chinese researchers first reported the use of these materials as orbital implants in the late 1990s.⁵² Glass-ceramic porous spheres of unspecified composition were implanted in the orbital cavity of enucleated rabbits and no material rejection was observed over a 6-month follow-up. Mid-term ultrasound analysis at 3 months revealed implant vascularization, which reached 90% of the porous volume of the implant after 6 months. Fibrovascular reaction occurred in these glass-ceramic implants at a quicker rate compared to porous polyethylene, according to the results reported elsewhere in rabbits.¹¹

The same glass-ceramic spheres were then implanted in 102 enucleated human patients who apparently reported no material-related complications after a follow-up period ranging from 6 to 24 months.⁵³ Four patients experienced

postoperative complications attributable to the operative techniques as their conjunctiva was damaged during the removal of the stitches, and one implant needed to be substituted with a new one. All patients felt satisfied with their cosmetic appearance and ocular motility acquired, without the need for an additional procedure of implant pegging.

It is interesting to underline that these early studies were performed while the researchers were apparently unaware of the proangiogenic potential of bioactive glasses, the first evidence of which was published only some years later by Day et al (in vitro assessment using fibroblasts).^{54,55}

A sporadic but interesting application of bioactive glasses was reported by Heringer and Ng⁵⁶ who filled old pegged tracts of hydroxyapatite porous orbital implants in order to allow repegging. Specifically, the pegs and sleeves that were previously placed in the orbital implants of three patients were removed due to incorrect positioning (miscentering and radial deviation), which caused discomfort during the coupling with the ocular prosthesis. The tunnel was filled with glass particulate and, after 2 months, the implant was successfully drilled again to host a new titanium peg. No complications were reported in all patients over a 3-year follow-up and a satisfactory connection of the implant to the ocular prosthesis was achieved.

3.2 | 45S5 Bioglass[®]/polyethylene composite orbital implants

Bioactive glasses have been widely used for producing polymer-based biomedical composites over the past two decades.⁵⁷ Probably inspired by these previous studies, the researchers of Porex Surgical (Newman, GA, USA) explored the possibility of adding bioactive glass particles to porous polyethylene orbital implant (Medpor[®]). The line of Medpor[®] implants was launched in the 1980s and this porous polymer was produced by molding medical-grade high-density polyethylene particles into a spherical or conical shape with 30-70 vol% of porosity⁵⁸; they gained soon an increased popularity due to the lower cost compared to porous hydroxyapatite and alumina.⁵⁹ Mixing melt-derived 45S5 Bioglass[®] particles (Novabone[®]; NovaBone Products LLC, Alachua, FL, USA) throughout the Medpor[®] structure was thought as a promising mean to improve the fibrovascularization rate: hence, the resulting glass/polyethylene (30:70 volume ratio) composite product (tradename: Medpor[®] Plus[™] Sphere) was cleared for clinical use via the 510(k) process by the Food and Drug Administration in 2002 and, since then, has been marketed worldwide.

A relatively limited number of studies are available on this type of orbital implant. Choi et al⁶⁰ first investigated the effect of bioactive glass on the fibrovascularization of Medpor[®] Plus[™] Spheres in rabbits. Forty-eight animals were evenly divided into four groups according to the different

surgical techniques and implanted materials used: groups 3 and 4 received the Medpor[®] Plus[™] Sphere after enucleation or evisceration, respectively, while groups 1 and 2 received a glass-free Medpor[®] implant after the two surgical procedures (reference groups). Interestingly, histological examinations at 2 postoperative months revealed that there were no statistically significant differences among the four groups in terms of fibrovascular ingrowth. Hence, this early study suggested that, apparently, the presence of bioactive glass inclusions did not carry any added value for improving implant biointegration and *in vivo* outcomes.

Opposite results were obtained by Naik et al in a small clinical trial.⁶¹ Ten human patients underwent enucleation followed by implantation of glass/polyethylene composite spheres (five cases) or Medpor[®] ball (five cases). Magnetic resonance imaging analysis revealed a statistically significant increase in fibrovascularization rate, expressed as the percentage of tissue-filled pore volume at each time point, in the patients receiving the glass-containing implants compared to the Medpor[®] group (69 vs 58% at 1.5 months; 85 vs 76% at 4.5 months).

A more extensive clinical study was reported by Ma et al⁶² who reviewed the clinical outcomes of 170 human patients after placement of Medpor[®] Plus[™] Spheres following enucleation. Most patients (161 cases) experienced no complications (good motility of implant and ocular prosthesis, no cases of conjunctival thinning or inflammation), while excessive discharge and implant postoperative exposure occurred in two and seven cases, respectively; of those, eight patients

needed additional surgery. These results suggest that glass/polyethylene composite porous spheres may be a useful alternative to other options, but an actual clinical advantage remains unclear as a comparison with glass-free Medpor[®] or a reference implant was lacking in this study.

3.3 | Biosilicate[®]-derived implants

Around 2010, the Brazilian research group led by Profs. Zanotto and Peitl proposed the use of Biosilicate[®] (composition 23.75Na₂O-23.75CaO-48.5SiO₂-4P₂O₅ wt%) to make a new generation of glass-ceramic orbital implants to restore volume in the anophthalmic socket. The story of the concept and applications of Biosilicate[®] and its devitrified derivatives was recently reviewed by Crovace et al.⁶³ Initially developed to be an alternative to 45S5 Bioglass[®] for use in bone and dental repair,^{64,65} Biosilicate[®]-derived glass-ceramics were found to be active also in contact with soft tissues, which is key for orbital implants.

In a first study published in 2012, Brandão et al⁶⁶ assessed the biocompatibility of cones composed by Biosilicate[®] or 45S5 Bioglass[®] (Figure 3A) in the eviscerated right eye of male albino Norfolk rabbits. Cones were produced by casting the melt into graphite molds; no crystallization was induced in 45S5 Bioglass[®] cones, whereas Biosilicate[®] cones underwent two different thermal treatments to deliberately develop one or two crystalline phases. Specifically, Biosilicate[®] 1P cones were treated so as to contain only one crystalline phase (1Na₂O·2CaO·3SiO₂), with P₂O₅ remaining in solid solution,

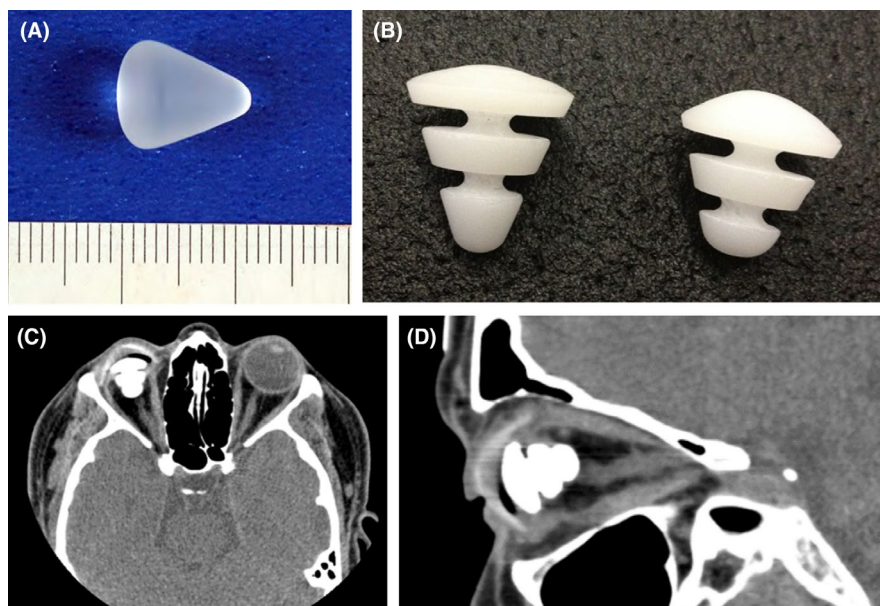


FIGURE 3 Biosilicate[®]-derived glass-ceramic orbital implants: (A) conical implant used in eviscerated rabbits; (B) tapered implants with circumferential channels used in human patients (two sizes available: left 18 mm, right 16 mm); (C) coronal and (D) sagittal tomographic imaging of a patient receiving a 16-mm long Biosilicate[®] tapered implant in the left orbit, with good positioning without migration and successful maintenance of the orbital volume after 180 days of follow-up. Images (A and B) courtesy of Oscar Peitl, (C and D) courtesy of Simone M. Brandão [Colour figure can be viewed at wileyonlinelibrary.com]

whereas the thermal treatment cycle chosen for Biosilicate[®] 2P allowed the phosphate ions to form an additional crystalline phase with calcium, thus creating apatite crystals. All cones were individually sterilized in ethylene oxide prior to use in vivo. The animals were divided into three groups that differed by type of conical biomaterial implanted, and were sacrificed at 7, 90, and 180 days after placement of the cones in the eviscerated scleral cavity. Over the whole follow-up period, none of the animals experienced orbital infection or implant migration/extrusion, and the morphological analyses revealed the formation of a fibrovascularized pseudocapsule around all the implants. The 45S5 Bioglass[®] and Biosilicate[®] 1P implants induced lower inflammation and less pseudocapsule formation compared to Biosilicate[®] 2P. The inflammatory reaction reached the maximum at 7 days after evisceration and cone placement, and then gradually diminished in all groups, especially in the 45S5 Bioglass[®] group. Similar results were obtained by the same research group in a second study carried out in 45 eviscerated rabbits.⁶⁷ On the basis of these animal studies, they concluded that Biosilicate[®] 1P could be a promising alternative to 45S5 Bioglass[®] for the management of the anophthalmic socket, as it elicited neither systemic nor local toxicity in the orbit of eviscerated rabbits.

Hence, an early clinical trial (intervention phase III prospective study) on this type of glass-ceramic was performed at two Brazilian University Hospitals (the Clinic Hospitals of the State University of São Paulo and University of São Paulo) from 2013 to 2016; the results of these studies in humans are shortly reported here for the first time. Forty-five patients were randomly recruited (with no differences in age, gender, and eye laterality) and separated according to the type of material implanted, that is, Biosilicate[®] 1P (received by two-thirds of patients) or PMMA (received by one-third of patients), used as a control. All implants were conic with identical design, available in two sizes (16 and 18 mm), and were manufactured individually by Prof. Oscar Peitl at the Laboratory of Vitreous Materials of the Federal University of São Carlos, São Paulo, Brazil (Figure 3B). Unlike the simple cones previously placed in rabbits,^{66,67} these tapered implants exhibited a new design with two circumferential channels promoting the physical attachment to soft orbital tissues and also biointegration. A proper milling/cutting/polishing equipment was designed and developed to guarantee the reproducibility of channel positioning and dimensions (depth and width) as well as surface finishing of all glass-ceramic implants. Clinical evaluations were performed preoperatively and at 7, 30, 60, 120, and 180 days after surgery. Systemic analyses, laboratory tests, and computed tomography (CT) of the orbits were performed preoperatively and 180 days after surgery. Thirty-eight patients completed the whole follow-up of 180 postsurgical days: both Biosilicate[®] 1P and PMMA conical implants resulted in a good clinical outcome, with no

significant infectious or inflammatory processes. Only one patient from the PMMA group experienced early extrusion of the implant, and another one from the same group had conjunctival dehiscence, which was spontaneously solved; both problems were supposed to be related to the operative technique rather than the type of material implanted. CT analyses showed no migration of the implants of both materials in all examined patients over the follow-up period (Figures 3C,D), and laboratory analyses revealed no damage or apparent alteration in vital organs associated to the ionic dissolution products released from Biosilicate[®] 1P.

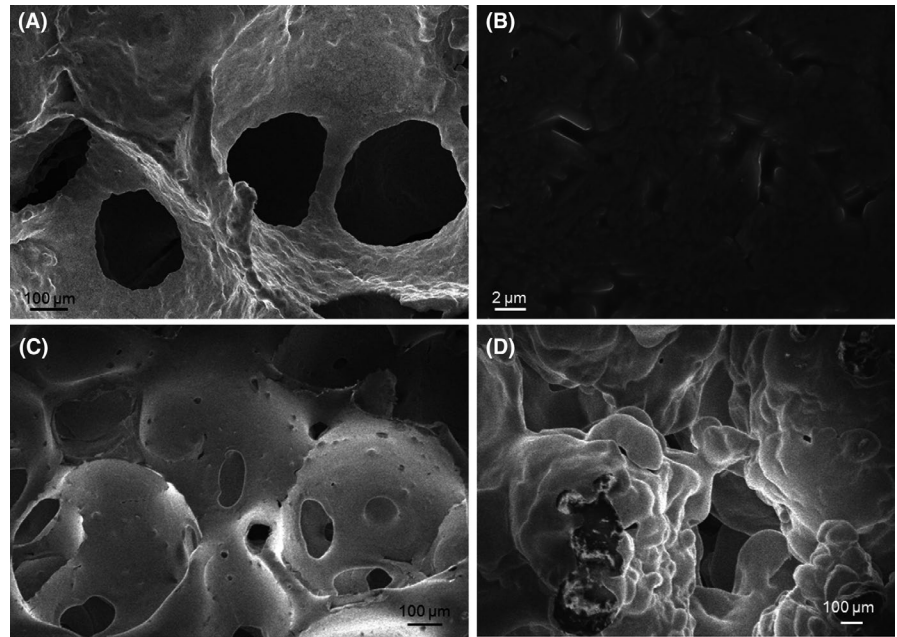
It is interesting to underline that, although not exhibiting an interconnected network of macropores, facilitating fibrovascular ingrowth, Biosilicate[®] tapered implants showed great promise from a clinical viewpoint due to the excellent biocompatibility, bactericidal activity—which positively contributes to minimizing postoperative infections—and overall positive biological response around the implant and in the orbital tissues. The relationship between this highly favorable behavior and the ionic dissolution products released by the material, as well as the impact of these ions on angiogenesis and typical pathogens involved in ocular infections, deserve to be further elucidated in future studies.

4 | CHALLENGES AND PERSPECTIVES

4.1 | Optimizing pore features/surface roughness and implant selection

The presence of an interconnected network of open macropores or channels in orbital implants inherently promotes fibrovascularization, which was reported to occur faster in ceramic implants compared to the relatively cheap polyethylene due to the more favorable surface chemistry for tissue ingrowth.¹¹ In the search for a less-expensive nonpolymeric alternative to macroporous-sintered alumina, sponge replication has been recently proved to be a highly promising method to fabricate glass-ceramic porous orbital implants due to easiness of execution, low cost and high versatility.⁶⁸ A nearly inert glass composition (57SiO₂-30CaO-6Na₂O-7Al₂O₃ mol%) was used to manufacture CaSiO₃-containing foams having a network of open pores (total porosity 55 vol.%, mean pore size 240 μm) potentially available for fibrovascular tissue ingrowth (Figures 4A,B). These implants were enough strong (compressive strength 20 MPa) to permit safe manipulation during surgery as well as postoperative integrity, which was also assured by excellent chemical stability in contact with biological fluids. This study pointed out that sponge-replicated glass-ceramic implants exhibited a similar pore-strut architecture compared to alumina implants (Figure 4C), but were markedly different from porous polyethylene implant (Figure 4D).

FIGURE 4 SEM micrographs showing the porous structure of (A) experimental glass-ceramic implant (composition: 57SiO₂-30CaO-6Na₂O-7Al₂O₃ mol%) with (B) detail of the surface showing CaSiO₃ crystals, (C) alumina implant, and (D) Medpor[®] sphere. The glass-ceramic and alumina implants exhibit a typical foam-like architecture, whereas the porous polyethylene has irregular pores with nonuniform and irregular struts. Images reproduced from Ref. ⁶⁸



Other silicate glass-ceramic porous implants of similar composition (57SiO₂-34CaO-6Na₂O-3Al₂O₃ mol% [SCNA]) were also fabricated by foam replication.⁶⁹ Very interestingly, the surface roughness (R_a) of these SCNA-based implants measured by contact profilometry was 2.5 times lower than that of porous alumina (300 vs 750 nm), which to date is considered as the “gold standard” option by many ophthalmic surgeons. Atomic force microscopy carried out on the same samples confirmed these early results and, furthermore, suggested that SCNA-derived glass-ceramic implants have comparable surface roughness to porous polyethylene, too.⁷⁰ A similar trend of the ranges of surface roughness was also found by examining foam-replicated glass-ceramic orbital implants based on a six-oxide glass composition (45SiO₂-26CaO-15Na₂O-7MgO-4K₂O-3P₂O₅ mol% [CEL2]).⁶⁹ From a clinical viewpoint, this is a potentially very important achievement since lower the surface roughness, lower the risk of conjunctival abrasion *in vivo*, and better the postoperative performance of a given implant.

Although favorable surface roughness and pore characteristics can indeed support the suitability of glass-ceramic materials (in this case, SCNA and CEL2) for making porous implants, a more robust approach is needed to reliably compare them to the other available solutions. When those studies were published,^{68–70} a widely accepted criterion to “globally” compare the structures and topographical characteristics of orbital implants did not exist. In order to bridge the gap, following a conceptualization previously adopted to compare tissue engineering scaffolds with spongy bone,^{71,72} Baino et al⁷³ tackled the challenge of developing an objective and quantitative approach for scoring porous orbital implant materials with different microstructural

characteristics. In order to compare the microarchitecture of pairs of implants (eg, porous glass-ceramic vs alumina or polyethylene), a multiparametric orbital implant similarity score (OrbISS) was defined as the squared distance between the materials in the six-dimensional space of the six selected key features, that is, total porosity, pore interconnectivity, specific surface area, pore connectivity density, degree of anisotropy, surface roughness—which were all previously assessed by micro-CT—and surface profilometry. According to its definition, the smaller this “global” index, the more similar the two samples of the pair. It was assessed that SCNA- and CEL2-derived glass-ceramic implants were similar to each other and to the alumina implant, while all ceramic implants were highly different from the porous polyethylene. These similarities and differences (eg, in terms of pore size/shape and strut thickness) can be roughly seen by visual inspection of micro-CT reconstructions (Figure 5) and confirms previous SEM observations⁶⁸ (see Figure 4). This approach can be easily extended to quantify how new glass and glass-ceramic porous implants are morphologically “distant” from reference (commercial) implants.

This similarity index could also be exploited for predictive purposes, as the clinical performance of orbital implants strongly depends on the material-related parameters included in OrbISS. Its use could make the selection of orbital implants less arbitrary and less dependent on the skills and personal preference of ophthalmic surgeons. Future studies should be addressed to improve the prediction capability of OrbISS by incorporating additional parameters not limited to implant architecture, such as ion dissolution kinetics—if relevant—and therapeutic effects of ions, and appropriate weights for each parameter, thus taking into account the relative importance

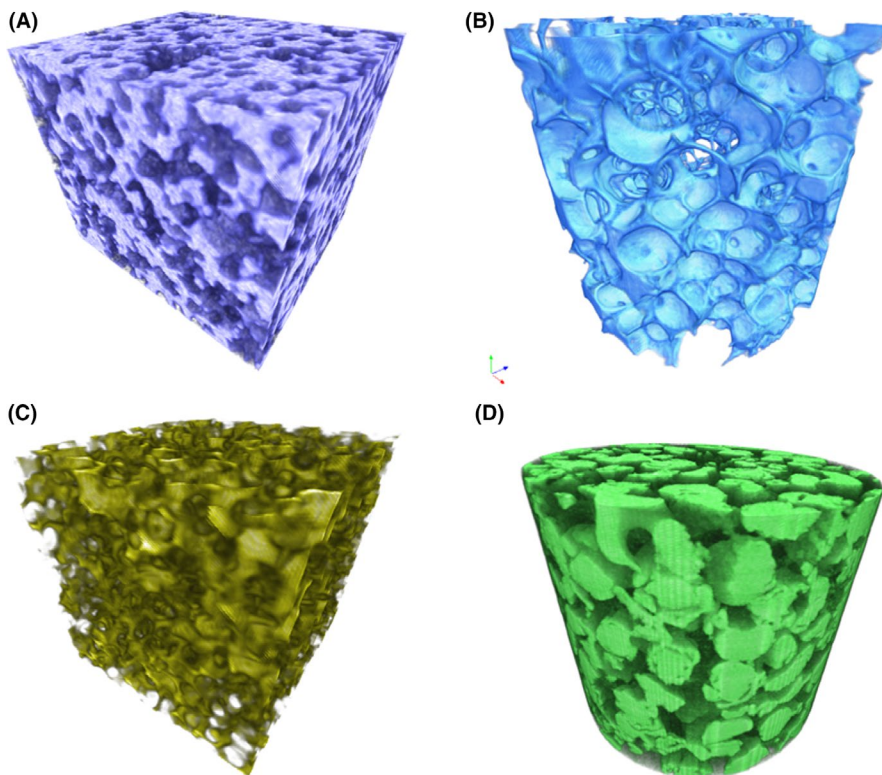


FIGURE 5 3D microtomographic reconstructions of representative subvolumes of different orbital implants: (A) SCNA-derived glass-ceramic (glass composition: 57SiO₂-34CaO-6Na₂O-3Al₂O₃ mol%), (B) CEL2-based glass-ceramic (glass composition: 45SiO₂-26CaO-15Na₂O-7MgO-4K₂O-3P₂O₅ mol%), (C) alumina, (D) Medpor[®] sphere. Images reproduced from Ref. ⁷³ [Colour figure can be viewed at wileyonlinelibrary.com]

correlated with clinical performance. In principle, the concepts behind the original OrbISS, developed for foam-like monophasic implants, can be extended to other pore/wall geometries and even multimaterial formulations.

Additive manufacturing (AM) of bioactive materials and composites has recently gained increasing interest in the biomedical community and is currently regarded as the last frontier of medical implant fabrication as it allows an accurate design and control of their internal structure.⁷⁴ AM-based approaches have been widely employed to develop tissue engineering bioactive glass scaffolds for the repair of bone and osteochondral defects⁷⁵; however, their applicability in ophthalmology is still limited to few studies addressed to orbital floor repair (45S5 Bioglass[®] porous meshes produced by stereolithography⁷⁶ or laser-cladded nonporous plates⁷⁷). At present, AM in the field of orbital implants has not been experimented yet, albeit carrying an enormous potential; this gap deserves to be bridged in the next few years.

4.2 | Antibacterial properties

Orbital implant infections, which are usually contracted as a result of implant exposure and colonization by bacteria, can be effectively treated by systemic antibiotics—if the implant is vascularized—or local therapy. Implant removal is the most drastic remedy that is carried out if the infection does not resolve pharmacologically, thereby implying additional cost and stress to the patient.^{78–80} Ophthalmic surgeons use to dip porous orbital implants in an antibiotic solution prior to

implantation in the orbital cavity.⁸¹ Although this approach is useful intraoperatively, it is ineffective in the long term to combat late or exposure-related infections. Furthermore, the abuse of antibiotics over the last decades has led to the development of resistant bacterial strains,⁸² which are a global challenge for the 21st century and need to be treated by following different approaches.

At present, neither commercial orbital implants nor external ocular prostheses provided with inherent antiseptic properties are available on the market and, in general, there is a paucity of studies in this field. An acrylic ocular prosthesis embedding small amounts of silver nanoparticles throughout its volume (300–700 ppm) has been patented and proved to be effective against various bacterial strains *in vitro* (*Streptococcus pneumoniae*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, and *Escherichia coli*),^{83,84} but did not reach clinical applications yet. Another patented strategy involves the sputter deposition of an antibacterial layer on the walls of orbital implants and on the rear surface of acrylic ocular prostheses (ie, the area in contact with the conjunctiva).⁸⁵ This coating is made of silver nanoclusters (10–50 nm) embedded in a pure-silica glass matrix and is highly stable from chemical and mechanical viewpoints under dry conditions up to 500°C.⁸⁶ Upon soaking in biological fluids, the coating tends to progressively solubilize over time releasing silver ions that exert a potent antibacterial effect for above 1 month *in vitro*.⁸⁷ The antibacterial effect of silver ions (Ag⁺) is associated with the strong binding of silver with disulfide

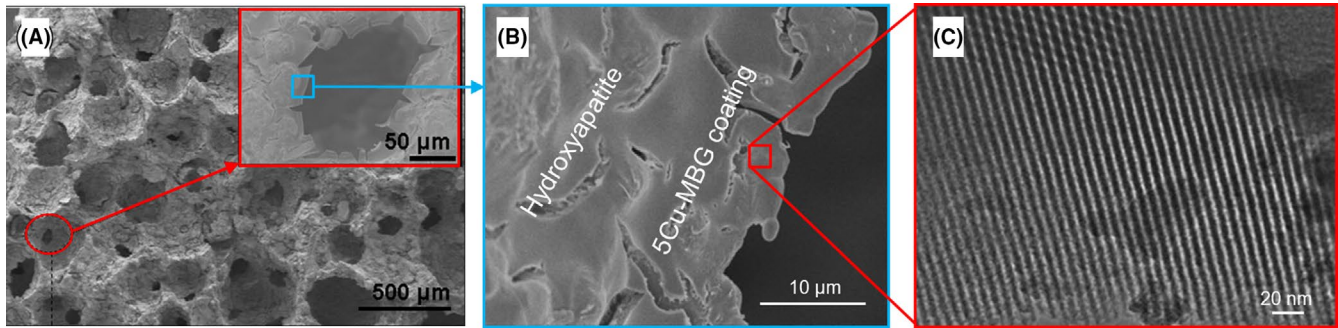


FIGURE 6 Mesoporous bioactive glass (MBG)-coated porous hydroxyapatite orbital implants: (A) overview of the porous surface, (B) interface between hydroxyapatite and Cu-doped MBG coating, (C) typical mesoporous texture (assessed by high-resolution TEM) of the MBG layer. Images adapted from Ref. ⁹¹ [Colour figure can be viewed at wileyonlinelibrary.com]

(S-S) and sulfhydryl (-SH) groups located on the proteins of microbial cell walls. After bonding with silver, the metabolic processes of bacteria (eg, oxidative metabolism and uptake of nutrients) stop, thereby leading to cell death.⁸⁸ This is a key advantage over the systems that release silver nanoparticles instead of ions, as the formers are associated to both acute and long-term toxicity and pose critical safety issues.⁸⁹ The intensity and duration of the antibacterial effect elicited by the sputter-deposited layer can be tailored by modulating the silver concentration (through acting on the deposition parameters like power and pressure in the sputtering chamber), the metal nanocluster size (which increases if post-sputtering thermal treatments are applied within 500–600°C), and the coating thickness (from tens of nanometers to few micrometers).⁹⁰

A conceptually similar approach, based on the deposition of an antibacterial glass-based surface layer, was reported by Ye et al⁹¹ who coated porous hydroxyapatite orbital implants with a Cu-doped mesoporous bioactive glass (MBG). MBGs are generally produced by incorporating supramolecular chemistry (evaporation-induced self-assembly) in the sol-gel method and are well recognized as versatile platforms for the controlled release of a number of drugs and therapeutic ions.⁹² The aim of that study was to synergistically combine the antibacterial effects of released copper ions, which are able to kill bacteria via the generation of reactive oxygen species, lipid peroxidation, protein oxidation, and DNA degradation,⁹³ and ofloxacin, an antibiotic hosted inside the mesopores (diameter from 3 to 5 nm). MBG coatings doped with 2 or 5 mol% of CuO were deposited by dipping of the hydroxyapatite implant in the sol and then consolidated via thermal treatment (Figure 6). In vitro tests showed that both Cu-doped implants inhibited the viability of *Staphylococcus aureus* and *E. coli*; the antibacterial halo increased from about 12 to 15 mm as the copper content increased, although the drug loading and release capacity was less efficient in the samples with higher copper concentration. This trend apparently suggests a predominant antiseptic effect associated to the release of copper ions.

It is worth highlighting that the coating-based approaches are successful in fulfilling two apparently irreconcilable requirements of these orbital implants, that is, the need for a permanent material (the skeleton lying underneath the coating) combined with the release of antibacterial ions (silver, copper) that occurs as the glass layer dissolves over time.

Another aspect also deserves to be highlighted: some bioactive glass compositions were shown to elicit an inherent antiseptic activity, without the need for doping with specific metallic cations, due to the local increment of pH associated to the release of alkaline ions (primarily Na⁺ and Ca²⁺) in the biological fluids. Perhaps the most famous example is represented by the S53P4 glass (53SiO₂–23Na₂O–20CaO–4P₂O₅ wt%), which is commercially sold as an antibacterial product (BoneAlive[®]; BoneAlive, Turku, Finland) for oral and dental applications. Stoor et al investigated the effect of S53P4 on a wide range of oral pathogens in a series of studies carried out in humans.^{94–96} S53P4 paste was reported to exhibit a potent and relatively fast antimicrobial effect (from 10 to 60 minutes depending on the type of bacteria) in inhibiting the viability of microorganisms of both supra- and subgingival plaques.⁹⁶ S53P4 granules and disks were also used as interpositional implants in 11 human patients suffering from nasal septum perforations; successful closure was obtained in 10 cases and no implant extrusions or infections in the nasal cavity were reported over 37 months of follow-up.⁹⁷ Good clinical outcomes were also obtained in the treatment of atrophic rhinitis associated to *Klebsiella ozaenae*.⁹⁶ Future investigations should address the antiseptic properties of S53P4 composition against the pathogens commonly associated to ocular infections and failure of orbital implants in order to assess its suitability for this new application. Furthermore, similar studies should be also performed on 45S5 Bioglass[®] and Biosilicate[®], which have already been successfully used for producing orbital implants; surprisingly, there is a lack of relevant reports in the literature on this specific point.

To combat infections through the action of antiseptic metal ions released from implant surfaces is a valuable and promising strategy in many biomedical fields.⁹⁷ However, several peculiar parameters related to the “working conditions” of orbital implants and external ocular prostheses should be taken into account for designing clinically safe biomaterials: for example, the interaction of metal ions with ocular secretions, the fate of released ions, and the associated risk of local storage and tissue necrosis are all issues deserving careful consideration. An extreme case of corneal argyrosis was reported in a 67-year-old woman wearing silver nitrate-coated cosmetic soft contact lenses over 17 years for the treatment of diplopia⁹⁸: this is a typical example of how an unknown and unpredicted ion-related side effect may be revealed only after many years of follow-up.

4.3 | Improving fibrovascularization

Besides exerting an antibacterial effect, copper ions are known to regulate the expression of many factors involved in angiogenesis, such as VEGF, FGF1/2, fibronectin, angiogenin, collagenase, prostaglandin E-1, and ceruloplasmin, which have key roles in initiation (vasodilation and vascular permeabilization), maturation (endothelial cell proliferation, migration, and morphogenesis), and regulation of blood vessel formation.^{99,100} From a biomolecular viewpoint, copper-induced angiogenesis is thought to be related with the mitogen-activated protein kinase (MAPK) signaling pathway, leading to endothelial cell sprouting.¹⁰¹ This property can be helpful to accelerate wound healing, as shown in some animal studies (rat model).^{102–104} Based on this evidence, Bairo first suggested in 2015¹⁰⁵ that the Cu-doped MBG coating developed by Ye et al⁹¹ could be useful to promote fibrovascularization in porous orbital implants due to controlled delivery of copper ions. This hypothesis was actually verified in vivo in 2018 by Ye's group,¹⁰⁶ who performed primary angiogenic tests in a panniculus carnosus muscle model in rabbits and reported that the Cu-doped glass coating significantly accelerated the vascularization of porous hydroxyapatite orbital implants compared to Cu-free materials.

Incorporation of copper in glass-ceramic orbital implants was also reported using a nearly inert alumina-silicate glass as a base material.¹⁰⁷ In a first approach, melt-derived Cu-doped strong macroporous scaffolds (compressive strength about 20 MPa) were produced by sponge replication, but the release of copper ions was inadequate to elicit a therapeutic effect. The second strategy, involving the deposition of a thin Cu-doped MBG layer on the walls of the previously prepared porous glass-ceramic foam, allowed a more sustained release of copper to be achieved, thereby motivating further research on the biological suitability and therapeutic effects of these glass-based materials.

5 | CONCLUSIONS

Although bioactive glasses cannot recover the sight of an enucleated or eviscerated patient, they can indeed contribute to replace volume in the anophthalmic socket, improving the appearance and self-esteem, and facilitating the reintegration of the individual into society. Bioactive glasses can also improve the success of the surgical procedures and the performance of the orbital implants by imparting appealing added value and extra-functionalities. Through the local release of therapeutic ions (eg, copper), bioactive glasses carry the potential to accelerate implant vascularization, which is the key to ensure an adequate biointegration and motility of the orbital implant as well as a valuable mean to reduce the risk of postoperative infections. A direct antibacterial effect can be exerted by other ions embeddable in the glass implant, such as silver, and thus multifunctional implants provided with both antiseptic and proangiogenic properties could be obtained. Other advantages of using bioactive glasses include the low cost compared to sintered ceramic implants, which require higher processing temperatures, and tunable surface roughness that can be properly decreased to minimize the risk of conjunctival abrasion in vivo and implant exposure. Glasses are also relatively easy to manufacture in various porous or nonporous forms, and can be incorporated as bioactive inclusions in a polymeric soft matrix (Medpor[®] Plus[™] Sphere). The application of bioactive glass and glass-ceramic implants after enucleation or evisceration is yet in its beginning and is less popular compared to other “traditional” areas of application, like orthopedics and dentistry, but is expected to emerge in the next few years, thus further expanding the benefits of glass in medicine.

ORCID

Francesco Bairo  <https://orcid.org/0000-0001-8860-0497>

REFERENCES

1. Moshfeghi DM, Moshfeghi AA, Finger PT. Enucleation. *Surv Ophthalmol*. 2000;44:277–301.
2. Kelley JJ. History of ocular prostheses. *Int Ophthalmol Clin*. 1970;10:713–9.
3. Luce CM. A short history of enucleation. *Int Ophthalmol Clin*. 1970;10:681–7.
4. Tonkelaar ID, Henkes HE, Leersum GK. Herman Snellen (1834-1908) and Muller's 'Reform-Auge' - a short history of the artificial eye. *Doc Ophthalmol*. 1991;77:349–54.
5. Mules PH. Evisceration of the globe with artificial vitreous. *Trans Ophthalmol Soc UK*. 1885;5:200–6.
6. Bairo F, Potestio I. Orbital implants: state-of-the-art review with emphasis on biomaterials and recent advances. *Mater Sci Eng, C*. 2016;69:1410–28.

7. Danz W Sr. Ancient and contemporary history of artificial eyes. *Adv Ophthalm Plast Reconstr Surg.* 1990;8:1–10.
8. Perry AC. Advances in enucleation. *Ophthalm Clin North Am.* 1991;4:173–82.
9. Jordan DR, Stoica B, Klapper SR. Current indications for pegging in the anophthalmic socket: are there any? *Curr Opin Ophthalmol.* 2016;27:465–73.
10. Karesh JW. Biomaterials in ophthalmic plastic and reconstructive surgery. *Curr Opin Ophthalmol.* 1998;9:66–74.
11. Jordan DR, Brownstein S, Dorey M, Yuen VH, Gilberg S. Fibrovascularization of porous polyethylene (Medpor) orbital implant in a rabbit model. *Ophthalmic Plast Reconstr Surg.* 2004;20:136–43.
12. Baino F, Perero S, Ferraris S, Miola M, Balagna C, Verné E, et al. Biomaterials for orbital implants and ocular prostheses: overview and future prospects. *Acta Biomater.* 2014;10:1064–87.
13. Chalasani R, Poole-Warren L, Conway RM, Ben-Nissan B. Porous orbital implants in enucleation: a systematic review. *Surv Ophthalmol.* 2007;52:145–55.
14. Choi S, Lee SJ, Shin JH, Cheong Y, Lee HJ, Paek JH, et al. Ultrastructural investigation of intact orbital implant surfaces using atomic force microscopy. *Scanning.* 2011;33:211–21.
15. Mawn LA, Jordan DR, Gilberg S. Scanning electron microscopic examination of porous orbital implants. *Can J Ophthalmol.* 1998;33:203–9.
16. Xu LC, Siedlecki CA. Effects of surface wettability and contact time on protein adhesion to biomaterial surfaces. *Biomaterials.* 2007;28:3273–83.
17. Sousa RLF, Schellini SA, Zornoff DCM, Padovani CR. Conduitas para reparação da cavidade anoftálmica no Brasil. *Arq Bras Oftalmol.* 2012;75:394–7.
18. Ababneh OH, AboTaleb EA, Abu Ameerh MA, Yousef YA. Enucleation and evisceration at a tertiary care hospital in a developing country. *BMC Ophthalmol.* 2015;15:120.
19. Sami D, Young S, Petersen R. Perspective on orbital enucleation implants. *Surv Ophthalmol.* 2007;52:244–65.
20. Anderson RL, Thiese SM, Nerad JA, Jordan DR, Tse D, Allen L. The universal orbital implant: indications and methods. *Adv Ophthalmic Plast Reconstr Surg.* 1990;8:88–99.
21. Rubin PA, Popham J, Rumelt S, Remulla H, Bilyk JR, Holds J, et al. Enhancement of the cosmetic and functional outcome of enucleation with the conical orbital implant. *Ophthalmology.* 1998;105:919–25.
22. Mourits DL, Hartong DT, Bosscha MI, Kloos RJHM, Moll AC. Worldwide enucleation techniques and materials for treatment of retinoblastoma: an international survey. *PLoS ONE.* 2015;10:e0121292.
23. Mourits DL, Moll AC, Bosscha MI, Tan HS, Hartong DT. Orbital implants in retinoblastoma patients: 23 years of experience and a review of the literature. *Acta Ophthalmol.* 2016;94:165–74.
24. Bhattacharjee K, Bhattacharjee H, Kuri G, Das J, Dey D. Comparative analysis of use of porous orbital implant with mucus membrane graft and dermis fat graft as a primary procedure in reconstruction of severely contracted socket. *Indian J Ophthalmol.* 2014;62:145–53.
25. Ibanez-Flores N, Abia-Serrano M, Aznar-Pena I, Mascaro-Zamora F, Castellar-Cerpa J, Anaya-Alaminos R, et al. Pericranium grafts for exposed orbital implants: an observational case-series study. *J Craniomaxillofac Surg.* 2015;43:1017–20.
26. Guyton JS. Enucleation and allied procedures: a review and description of a new operation. *Trans Am Ophthalmol Soc.* 1948;46:472–527.
27. Culler AM. Orbital implants after enucleation: basic principles of anatomy and physiology of the orbit and relation to implant surgery. *Trans Am Acad Ophthalmol Otolaryngol.* 1952;56:17–20.
28. Colombo P. Conventional and novel processing methods for cellular ceramics. *Philos Trans A Math Phys Eng Sci.* 2006;364:109–24.
29. Hench LL. The story of Bioglass®. *J Mater Sci Mater Med.* 2006;17:967–78.
30. Hench LL. Genetic design of bioactive glasses. *J Eur Ceram Soc.* 2009;29:1257–65.
31. Jones JR, Brauer DS, Hupa L, Greenspan DC. Bioglass and bioactive glasses and their impact on healthcare. *Int J Appl Glass Sci.* 2016;7:423–34.
32. Montazerian M, Zanotto ED. Bioactive and inert dental glass-ceramics. *J Biomed Mater Res A.* 2017;105:619–39.
33. Hench LL, Greenspan D. Interactions between bioactive glass and collagen: a review and new perspectives. *J Aust Ceram Soc.* 2013;49:1–40.
34. Hench LL, Polak JM. Third-generation biomedical materials. *Science.* 2002;295:1014–7.
35. Sanders DM, Hench LL. Mechanisms of glass corrosion. *J Am Ceram Soc.* 1973;56:373–7.
36. Baino F. Bioactive glasses - when glass science and technology meet regenerative medicine. *Ceram Int.* 2018;44:14953–66.
37. Rahaman MN, Day DE, Bal BS, Fu Q, Jung SB, Bonewald LF, et al. Bioactive glass in tissue engineering. *Acta Biomater.* 2011;7:2355–73.
38. Baino F, Novajra G, Miguez-Pacheco V, Boccaccini AR, Vitale-Brovarone C. Bioactive glasses: special applications outside the skeletal system. *J Non-Cryst Solids.* 2016;432:15–30.
39. Kargozar S, Baino F, Hamzehlou S, Hill RG, Mozafari M. Bioactive glasses: sprouting angiogenesis in tissue engineering. *Trends Biotechnol.* 2018;36:430–44.
40. Gorustovich AA, Roether JA, Boccaccini AR. Effect of bioactive glasses on angiogenesis: a review of in vitro and in vivo evidences. *Tissue Eng B.* 2009;16:199–207.
41. Gerhardt LC, Widdows KL, Erol MM, Burch CW, Sanz-Herrera JA, Ochoa I, et al. The pro-angiogenic properties of multifunctional bioactive glass composite scaffolds. *Biomaterials.* 2011;32:4096–108.
42. Li H, Xue K, Kong N, Liu K, Chang J. Silicate bioceramics enhanced vascularization and osteogenesis through stimulating interactions between endothelial cells and bone marrow stromal cells. *Biomaterials.* 2014;35:3803–18.
43. Munaron L. Intracellular calcium, endothelial cells and angiogenesis. *Recent Pat Anticancer Drug Discov.* 2006;1:105–19.
44. Bose S, Fielding G, Tarafder S, Bandyopadhyay A. Understanding of dopant-induced osteogenesis and angiogenesis in calcium phosphate ceramics. *Trends Biotechnol.* 2013;31:594–605.
45. Mao C, Chen X, Miao G, Lin C. Angiogenesis stimulated by novel nanoscale bioactive glasses. *Biomed Mater.* 2015;10:025005.
46. Wray P. “Cotton candy” that heals. *Am Ceram Soc Bull.* 2011;90:24–31.
47. Kaur G, Pandey OP, Singh K, Homa D, Scott B, Pickrell GA. Review of bioactive glasses: their structure, properties, fabrication and apatite formation. *J Biomed Mater Res A.* 2014;102:254–74.

48. Montazerian M, Zanotto ED. History and trends of bioactive glass-ceramics. *J Biomed Mater Res A*. 2016;104:1231–49.
49. Baino F, Verné E. Glass-based coatings on biomedical implants: a state-of-the-art review. *Biomed Glasses*. 2017;3:1–17.
50. Cacciotti I. Bivalent cationic ions doped bioactive glasses: the influence of magnesium, zinc, strontium and copper on the physical and biological properties. *J Mater Sci*. 2017;52:8812–31.
51. El-Rashidy AA, Roether JA, Harhaus L, Kneser U, Boccaccini AR. Regenerating bone with bioactive glass scaffolds: a review of in vivo studies in bone defect models. *Acta Biomater*. 2017;62:1–28.
52. Xu X, Wang C, Huang T, Ding L, Huang Z, Zhang X. An experimental study of bioactive glass ceramics as orbital implants. *Bull Hunan Med Univ*. 1997;22:25–8.
53. Xu X, Huang Z, Wang C. Clinical study of bioactive glass ceramics as orbital implants. *Bull Hunan Med Univ*. 1997;22:440–2.
54. Day RM. Bioactive glass stimulates the secretion of angiogenic growth factors and angiogenesis in vitro. *Tissue Eng*. 2005;11:768–77.
55. Day RM, Maquet V, Boccaccini AR, Jerome R, Forbes A. In vitro and in vivo analysis of macroporous biodegradable poly(D,L-lactide-co-glycolide) scaffolds containing bioactive glass. *J Biomed Mater Res A*. 2005;75:778–87.
56. Heringer DM, Ng JD. A novel approach to re-pegging hydroxyapatite implants using bioactive glass. *Ophthalmic Plast Reconstr Surg*. 2006;22:45–7.
57. Rezwani K, Chen QZ, Blaker JJ, Boccaccini AR. Biodegradable and bioactive porous polymer/inorganic composite scaffolds for bone tissue engineering. *Biomaterials*. 2006;27:3413–31.
58. Karesh JW, Dresner SC, Dutton JJ. High-density porous polyethylene (Medpor) as a successful anophthalmic socket implant. *Ophthalmology*. 1994;101:1688–96.
59. Jung S, Cho W, Paik J, Yang S. Long-term surgical outcomes of porous polyethylene orbital implants: a review of 314 cases. *Br J Ophthalmol*. 2012;96:494–8.
60. Choi HY, Lee JE, Park HJ, Oum BS. Effect of synthetic bone glass particulate on the fibrovascularization of porous polyethylene orbital implants. *Ophthalmic Plast Reconstr Surg*. 2006;22:121–5.
61. Naik MN, Murthy RK, Honavar SG. Comparison of vascularization of Medpor and Medpor-plus orbital implants: a prospective, randomized study. *Ophthalmic Plast Reconstr Surg*. 2007;6:463–7.
62. Ma X, Schou KR, Maloney-Schou M, Harwin FM, Ng JD. The porous polyethylene/bioglass spherical orbital implant: a retrospective study of 170 cases. *Ophthalmic Plast Reconstr Surg*. 2011;27:21–7.
63. Crovace MC, Souza MT, Chinaglia CR, Peitl O, Zanotto ED. Biosilicate® - a multipurpose, highly bioactive glass-ceramic. In vitro, in vivo and clinical trials. *J Non-Cryst Solids*. 2016;432:90–110.
64. Peitl O, Zanotto ED, Hench LL. Bioactive ceramics and method for preparing bioactive ceramics. Patent WO 97/41079, 1997.
65. Zanotto ED, Ravagnani C, Peitl O, Panzeri H, Lara EG. Process and compositions for preparing particulate, bioactive or resorbable biosilicates for use in the treatment of oral ailments. Patent WO 2004074199 A1, 2007.
66. Brandao SM, Schellini SA, Moraes AD, Padovani CR, Pellizzon CH, Peitl O, et al. Biocompatibility analysis of Bioglass 45S5 and Biosilicate implants in the rabbit eviscerated socket. *Orbit*. 2012;31:143–9.
67. Brandao SM, Schellini SA, Padovani CR, Peitl O, Hashimoto E. Biocompatibility analysis of Bioglass 45S5 and Biosilicate cone in rabbit eviscerated cavity. *Rev Bras Oftalmol*. 2013;72:21–5.
68. Baino F. Porous glass-ceramic orbital implants: a feasibility study. *Mater Lett*. 2018;212:12–5.
69. Baino F, Gautier di Confienzo G, Faga MG. Fabrication and morphological characterization of glass-ceramic orbital implants. *Int J Appl Ceramic Technol*. 2018;15:884–91.
70. Salerno M, Reverberi A, Baino F. Nanoscale topographical characterization of orbital implant materials. *Materials*. 2018;11:660.
71. Falvo D'Urso Labate G, Baino F, Terzini M, Audenino AL, Vitale-Brovarene C, Segers P, et al. Bone structural similarity score: a multiparametric tool to match properties of biomimetic bone substitutes with their target tissues. *J Appl Biomater Funct Mater*. 2016;14:e277–89.
72. Falvo D'Urso Labate G, Catapano G, Vitale-Brovarene C, Baino F. Quantifying the micro-architectural similarity of bioceramic scaffolds to bone. *Ceram Int*. 2017;43:9443–50.
73. Baino F, Falvo D'Urso Labate G, Gautier di Confienzo G, Faga MG, Vitale-Brovarene C, Catapano G. Microstructural characterization and robust comparison of ceramic porous orbital implants. *J Eur Ceram Soc*. 2018;38:2988–93.
74. Singh S, Ramakrishna S. Biomedical applications of additive manufacturing: present and future. *Curr Opin Biomed Eng*. 2017;2:105–15.
75. Gmeiner R, Deisinger U, Schönherr J, Lechner B, Detsch R, Boccaccini AR, et al. Additive manufacturing of bioactive glasses and silicate bioceramics. *J Ceram Sci Technol*. 2015;6:75–86.
76. Tesavibul P, Felzmann R, Gruber S, Liska R, Thompson I, Boccaccini AR, et al. Processing of 45S5 Bioglass® by lithography-based additive manufacturing. *Mater Lett*. 2012;41:81–4.
77. Comesana R, Lusquiños F, Del Val J, Lopez-Álvarez M, Quintero F, Riveiro A, et al. Three-dimensional bioactive glass implants fabricated by rapid prototyping based on CO₂ laser cladding. *Acta Biomater*. 2011;7:3476–87.
78. You JR, Seo JH, Kim YH, Choi WC. Six cases of bacterial infection in porous orbital implants. *Jpn J Ophthalmol*. 2003;47:512–8.
79. Karsloglu S, Serin D, Simşek I, Ziyilan S. Implant infection in porous orbital implants. *Ophthalmic Plast Reconstr Surg*. 2006;22:461–6.
80. Jordan DR, Brownstein S, Robinson J. Infected aluminum oxide orbital implant. *Ophthalmic Plast Reconstr Surg*. 2006;22:66–7.
81. Badilla J, Dolman PJ. Methods of antibiotic instillation in porous orbital implants. *Ophthalmic Plast Reconstr Surg*. 2008;24:287–9.
82. Arias CA, Murray BE. Antibiotic-resistant bugs in the 21st century - a clinical super-challenge. *N Engl J Med*. 2009;360:439–43.
83. Jun MS, Jun JH, Jun SU, Jun YM. Bio-artificial eye and conformer. US Patent No. US2008/0262612A1; 2008.
84. Yang JW, Choi JW, Lee SG, Kim DS. Antibacterial properties of artificial eyes containing nano-sized particle silver. *Orbit*. 2011;30:77–81.
85. Baino F, Perero S, Miola M, Ferraris S, Verne E, Ferraris M. Rivestimenti e trattamenti superficiali per impartire proprietà antibatteriche a dispositivi per oftalmoplastica. IT Patent No. TO2012A00051; 2012.

86. Ferraris M, Perero S, Miola M, Ferraris S, Verne E, Morgiel J. Silver nanocluster–silica composite coatings with antibacterial properties. *Mater Chem Phys*. 2010;120:123–6.
87. Bairo F, Ferraris S, Miola M, Perero S, Verné E, Coggiola A, et al. Novel antibacterial ocular prostheses: proof of concept and physico-chemical characterization. *Mater Sci Eng, C*. 2016;60:467–74.
88. Jung WK, Koo HC, Kim KW, Shin S, Kim SH, Park YH. Antibacterial activity and mechanism of action of the silver ion in *Staphylococcus aureus* and *Escherichia coli*. *Appl Environ Microbiol*. 2008;74:2171–8.
89. Caballero-Diaz E, Pfeiffer C, Kastl L, Rivera-Gil P, Simonet B, Valcarcel M, et al. The toxicity of silver nanoparticles depends on their uptake by cells and thus on their surface chemistry. *Part Part Syst Charact*. 2013;30:1079–85.
90. Ferraris M, Balagna C, Perero S, Miola M, Ferraris S, Bairo F, et al. Silver nanocluster/silica composite coatings obtained by sputtering for antibacterial applications. *IOP Conf Series Mater Sci Eng*. 2012;40:012037.
91. Ye J, He J, Wang C, Yao K, Gou Z. Copper-containing mesoporous bioactive glass coatings on orbital implants for improving drug delivery capacity and antibacterial activity. *Biotechnol Lett*. 2014;36:961–8.
92. Wu C, Chang J. Multifunctional mesoporous bioactive glasses for effective delivery of therapeutic ions and drug/growth factors. *J Controll Rel*. 2014;193:282–95.
93. Chatterjee AK, Chakraborty R, Basu T. Mechanism of antibacterial activity of copper nanoparticles. *Nanotechnology*. 2014;25:135101.
94. Stoor P, Soderling E, Salonen JJ. Antibacterial effects of a bioactive glass paste on oral microorganisms. *Acta Odontol Scand*. 1998;56:161–5.
95. Stoor P, Soderling E, Grenman R. Bioactive glass S53P4 in repair of septal perforations and its interactions with the respiratory infection-associated microorganisms *Haemophilus influenzae* and *Streptococcus pneumoniae*. *J Biomed Mater Res*. 2001;58:113–20.
96. Stoor P, Soderling E, Grenman R. Interactions between the bioactive glass S53P4 and the atrophic rhinitis-associated microorganism *Klebsiella ozaenae*. *J Biomed Mater Res*. 1999;48:869–74.
97. Hoppe A, Guldal Boccaccini AR. A review of the biological response to ionic dissolution products from bioactive glasses and glass-ceramics. *Biomaterials*. 2011;32:2757–74.
98. Hau SC, Tuft SJ. Presumed corneal argyrosis from occlusive soft contact lenses: a case report. *Cornea*. 2009;28:703–5.
99. Giacomelli C, Trincavelli ML, Satriano C, Hansson O, La Mendola D, Rizzarelli E, et al. Copper (II) ions modulate angiogenin activity in human endothelial cells. *Int J Biochem Cell Biol*. 2015;60:185–96.
100. Urso E, Maffia M. Behind the link between copper and angiogenesis: established mechanisms and an overview on the role of vascular copper transport systems. *J Vasc Res*. 2015;52:172–96.
101. Mavria G, Vercoulen Y, Yeo M, Paterson H, Karasandes M, Marais R, et al. ERK-MAPK signaling opposes Rho-kinase to promote endothelial cell survival and sprouting during angiogenesis. *Cancer Cell*. 2006;9:33–44.
102. Bi L, Rahaman MN, Day DE, Brown Z, Samujh C, Liu X, et al. Effect of bioactive borate glass microstructure on bone regeneration, angiogenesis, and hydroxyapatite conversion in a rat calvarial defect model. *Acta Biomater*. 2013;9:8015–26.
103. Zhao S, Li L, Wang H, Zhang Y, Cheng X, Zhou N, et al. Wound dressings composed of copper-doped borate bioactive glass microfibers stimulate angiogenesis and heal full-thickness skin defects in a rodent model. *Biomaterials*. 2015;53:379–91.
104. Bühner G, Rottensteiner U, Hoppe A, Detsch R, Dafinova D, Fey T, et al. Evaluation of in vivo angiogenic effects of copper doped bioactive glass scaffolds in the AV loop model. *Biomed Glasses*. 2016;2:111–7.
105. Bairo F. How can bioactive glasses be useful in ocular surgery? *J Biomed Mater Res A*. 2015;103:1259–75.
106. Wang C, Jin K, He J, Wang J, Yang X, Yao C, et al. Synergistic effect of copper-containing mesoporous bioactive glass coating on stimulating vascularization of porous hydroxyapatite orbital implants in rabbits. *J Biomed Nanotechnol*. 2018;14:688–97.
107. Bairo F, Potestio I, Vitale-Brovarone C. Production and physico-chemical characterization of Cu-doped silicate bioceramic scaffolds. *Materials*. 2018;11:1524.

How to cite this article: Bairo F, Verné E, Fiume E, et al. Bioactive glass and glass-ceramic orbital implants. *Int J Appl Ceram Technol*. 2019;16:1850–1863. <https://doi.org/10.1111/ijac.13236>